

1 **The Effects of morpholine pre-treated and carboxymethylated cellulose nanofibrils on**
2 **the properties of alginate-based hydrogels**

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10 **Abstract**

11 The effects of varying percentage loadings of morpholine pre-treated cellulose nanofibrils
12 (MCNF) and carboxymethylated cellulose nanofibrils (CMCNF) on the aqueous swelling,
13 compressive modulus and viscoelastic properties of calcium-ion-crosslinked alginate
14 hydrogels were investigated. In addition, the pore structure of hydrogels with the highest
15 compressive modulus were studied. The incorporation of 5 wt. % MCNF resulted in a slightly
16 reduced aqueous swelling, a 36 % increase in compressive modulus and a layered pore structure
17 when compared with the neat alginate hydrogel. On the other hand, the addition of CMCNF at
18 the same loading led to a slightly improved aqueous swelling, an increase in compressive
19 modulus (17 %) and high porosity. Further increases in CNF loadings did not result in
20 significant increase in material properties. The alginate/CNF composite materials have
21 potentials to be used in applications where good swelling and mechanical robustness are
22 required.

23 **Highlights**

- 24
- 25 • The effects of two types of CNF on alginate hydrogels were studied.
 - 26 • Aqueous swelling of the hydrogels was affected by the CNFs' surface properties and amounts.
 - 27 • Mechanical properties of the alginate hydrogels were improved by the addition of CNF.
 - 28 • The porosity of the hydrogels was influenced by the type of CNF.

29 *Keywords* Hydrogels, alginates, cellulose nanofibrils (CNF), carboxymethylation, surface
30 modification, compressive modulus.

31 **1. Introduction**

32 Polysaccharidic biopolymers such as alginates, cellulose, chitosan, chondroitin sulphate and
33 hyaluronic acid are widely researched for applications in biomedical (Aravamudhan, Ramos,
34 Nada, & Kumbar, 2014), pharmaceutical/drug delivery (Cardoso, Costa, & Mano, 2016),
35 packaging (Hubbe, Ferrer, Tyagi, Yin, & Salas, 2017), and agricultural fields (Guilherme et
36 al., 2015). These materials provide potentials for low cost and sustainable products because of
37 their ubiquitous sources of extraction. Moreover, their inherent non-toxic, biocompatible and
38 biodegradable properties have made them well recommended in biomedical applications for
39 bone and tissue regeneration (Park, Lih, Park, Joung, & Han, 2017), wound care dressings
40 (Leppiniemi et al., 2017) and for the delivery of various drugs and growth factors
41 (Aravamudhan et al., 2014; Augst, Kong, & Mooney, 2006).

42 Naturally occurring in seaweed and brown algae, Sodium Alginate (SA) is a linear
43 unbranched polymer consisting of β -D-mannuronic acid (M) and α -L-guluronic acid (G)
44 arranged in a chain sequence of MM-GG-MG (Augst et al., 2006). The presence of carboxyl
45 groups on the surface of SA promotes hydrophilicity and solubility in water. These carboxyl
46 groups can also engage in ionic crosslinking with divalent ions such as Ca^{2+} and Ba^{2+} in the
47 well-known “egg-box” model to form relatively stable network structures. The properties of
48 the resulting network structure are influenced by the type of crosslinking ions, concentration
49 of crosslinking ions, temperature, time and method of gelation (Kuo & Ma, 2001; Vicini,
50 Mauri, Wichert, & Castellano, 2017).

51 Calcium ions are mostly used for the ionic crosslinking of SA. These ions have been
52 introduced internally within the SA solution from a slurry of CaCO_3 -D-glucono- δ -lactone
53 system (Kuo & Ma, 2001), diffused from internally dispersed Ca^{2+} enriched microbeads
54 (Vicini, Castellano, Mauri, & Marsano, 2015; Vicini et al., 2017), diffused from a bath of CaCl_2
55 solution (Ma et al., 2017) or sprayed onto the SA solution (Lin et al., 2014). The internal
56 gelation method for SA crosslinking has been noted to provide more controlled gelation
57 compared to the external gelation/diffusion method, which usually results in a spontaneous
58 and heterogeneous material, characterised by a hard outer shell and a soft inner core (Kuo &
59 Ma, 2001). To reduce the rate of diffusion and gelation, Bajpai et al. (2016) added SA solution
60 to 8 kDa molecular weight cut off filtration membrane, before crosslinking in a bath of CaCl_2 .
61 The study shows that the gelation process started from the outer circumference of the SA

62 solution before gradually diffusing into the inner core. However, the homogeneity of the
63 resulting hydrogel prepared using this method was not investigated.

64 Regardless of the gelation method used for the crosslinking of sodium alginate, the use
65 of lower concentration of crosslinking ions leads to a material with high moisture absorbance
66 and low mechanical properties. Increasing the concentration of the crosslinking ions however
67 result to a material with low swelling and high mechanical properties (Augst et al., 2006). High
68 mechanical and good swelling properties are desirable properties in the fields of biomedicine.
69 In order to achieve the properties that would be fit for specific applications, many studies have
70 focused on formulating blends of alginate with synthetic and natural polymers (Kong &
71 Mooney, 2003; Lee et al., 2004; Lin et al., 2014). These alginate-based composites can be
72 formulated in the form of films, sponges and hydrogels (Augst et al., 2006). Being a three-
73 dimensional network material, hydrogels are capable of absorbing large amount of solvent
74 without dissolving and are therefore topical materials for various biomedical applications
75 (Ahmed, 2015).

76 Nanocellulose is a nano-sized cellulose material extracted in the form of rod-like
77 crystals (cellulose nanocrystals, CNC) and nanofibrils (cellulose nanofibrils, CNF) using
78 various chemical (Onyianta, Dorris, & Williams, 2018), enzymatic (Beyene et al., 2017) and
79 mechanical pre-treatments followed by mechanical processing (Abdul Khalil et al., 2014). It is
80 a renewable, biocompatible and biodegradable biopolymer with improved mechanical strength,
81 lightweight properties, optical properties, barrier properties and structuring capabilities
82 (Jonoobi et al., 2015; Klemm et al., 2011). Hence, the increased industrial and research interests
83 on nanocellulose for wide range of applications. Some of the chemical pre-treatment methods
84 such as carboxymethylation (Wågberg et al., 2008), 2,2,6,6-tetramethylpiperidine-1-oxyl
85 (TEMPO)-mediated oxidation (Isogai, Saito, & Fukuzumi, 2011), and quaternisation (Aulin,
86 Johansson, Wågberg, & Lindström, 2010) modify the surface of the cellulose, thereby
87 imparting new properties which are different from the original cellulose material. Onyianta et
88 al. (2018) compared the properties of cellulose nanofibrils prepared using three types of
89 chemical pre-treatment methods. The study shows that the presence or absence of surface
90 charge on the CNF is responsible for the changes in crystallinity, viscoelasticity, thermal
91 properties and aspect ratios of these CNF materials.

92 Interest in SA/nanocellulose blends in particular has increased, mainly because of the
93 reinforcing capability of nanocellulose (Huq et al., 2017; Lin et al., 2014; Ma et al., 2017;

94 Smyth et al., 2018). These blends of SA and nanocellulose are mostly prepared by external
95 gelation method, which is a fast gelling method with possible areas of localised gelation. In
96 addition, the carboxyl groups on the TEMPO-oxidised nanocellulose has been reported to
97 participate in the crosslinking process with Ca^{2+} ions (Lin, Bruzzese, & Dufresne, 2012).
98 However, studies on how the presence or absence of surface charge of the CNF and varying
99 amounts of CNF affect the elastically effective crosslinking chains of SA hydrogel are still
100 lacking.

101 In this study, slower diffusion and homogeneous crosslinking of SA hydrogels were
102 prepared using calcium chloride enriched agar wells, a method adapted from Stagnaro, Schizzi,
103 Utzeri, Marsano, & Castellano (2018). The effect of temperature and duration of crosslinking
104 on the homogeneity of the neat alginate hydrogels were initially studied. Subsequently, the
105 effects of unmodified and surface modified cellulose nanofibrils on the surface interaction,
106 equilibrium swelling, storage modulus, compressive modulus and morphology of alginate
107 hydrogels were investigated. The modified Flory's equation (Flory, 1953) for the deformation
108 of a swollen polymer network, based on the affine network model was then used to calculate
109 the effect of the two types of CNF on the elastically effective chains of SA hydrogels at varying
110 loadings. The affine network model assumes that macromolecular deformation of a network
111 structure directly translates to the deformation of the elastically effective chains within the
112 structure. This model has been used to characterise the elastically effective crosslinking degree
113 of other hydrogel materials from natural and synthetic origins (Muniz & Geuskens, 2001;
114 Sannino et al., 2005).

115 **2. Experimental**

116 **2.1 Materials**

117 Medium viscosity SA from brown algae, with molecular weight ranging from 80,000 to
118 120,000 g/mol and composed of approximately 61% mannuronic and 39% guluronic acids
119 (M/G ratio of 1.56), calcium chloride (CaCl_2) and sodium chloride (NaCl) were used as
120 received from Sigma-Aldrich. Agar powder by BD DifcoTM was used for the gel moulds. Two
121 types of aqueous CNF suspensions (1 wt. %) were used to prepare SA_CNF dispersions at
122 different loadings relative to the amount of SA solid content. The first CNF was produced using
123 aqueous morpholine pre-treatment before the mechanical processing (MCNF) and has
124 negligible surface charge. The second CNF was prepared by carboxymethylation pre-treatment

125 before the mechanical processing (CMCNF) and has 550 $\mu\text{mol/g}$ of anionic carboxymethyl
126 groups on its surface. The details and material properties of the two types of CNF used in this
127 study are well described previously by Onyianta et al. (2018). Deionised water was used
128 throughout the experiment.

129 **2.2 Preparation of SA solution and SA_CNF dispersions**

130 2 wt. % SA solution was prepared as a control by dissolving the appropriate amount of SA
131 powder in water under continuous stirring for not less than 2h using a magnetic stirrer. 100 g
132 SA_CNF dispersions were prepared by weighing out appropriate amounts of CNF suspension
133 into a beaker and diluted with the required amount of water and stirred. A constant mass of 2
134 g SA powder was then added to the diluted CNF suspension to make 2 wt. % SA having 5, 10
135 and 20 wt. % MCNF or CMCNF relative to the dry mass of SA, designated as SA_MCNF_5,
136 SA_MCNF_10, SA_MCNF_20, SA_CMCNF_5, SA_CMCNF_10 and SA_CMCNF_20. The
137 increment in CNF loading was by multiples of 2.

138 **2.3 Preparation of agar gel moulds and SA_CNF hydrogels**

139 Agar powder was added to 0.5 M CaCl_2 solution at 1 wt. % and brought to boil to dissolve the
140 agar. 15 g of the solution was weighed into small 55 x 15 mm petri-dishes and allowed to gel
141 at room temperature for not less than 2 h. After complete gelation of the agar solution,
142 cylindrical glasscutter with 28 mm internal diameter was used to create wells within the agar
143 gels. 4.5 g of SA solution or SA_CNF dispersions were then added into the wells created within
144 the agar gels. A rectangular glass slab was gently placed on top of the SA solution to prevent
145 bulging of the gels during crosslinking and to maintain the cylindrical shape of the hydrogels.
146 At the end of the crosslinking process, which was visually assessed by a change from
147 translucent light brownish colour to opaque light brownish colour, the gels were thoroughly
148 washed in deionised water and swollen to an equilibrium state.

149 **2.4 Equilibrium swelling of the never-dried hydrogels in water**

150 The never-dried hydrogels were swollen in water to an equilibrium point, with the aim of
151 characterising the network structure of the hydrogels. The mass of the equilibrium swollen
152 hydrogel and the oven dried mass were recorded and used for the calculation of the swelling
153 degree using the relationship given in Equation 1.

154 Swelling degree = $\left(\frac{W_s - W_d}{W_d}\right)$ Equation 1

155 Here, W_d is the dry mass of the hydrogel and W_s is the swollen mass of the hydrogel.

156 **2.5 Homogeneity Test**

157 A facile homogeneity test (Kuo & Ma, 2001) was initially carried out on the equilibrium
158 swollen neat SA hydrogels to study the effect of the crosslinking temperature and duration on
159 the uniformity of the crosslinks formed within the hydrogels. the tests were carried out at 4 °C,
160 25 °C and 37 °C for 24h, 48h and 72h. Since the crosslinking process occurred by diffusion of
161 the calcium ions from the agar gel into the SA solution, it is therefore expected that the outer
162 circumference of the SA solution in the mould would be crosslinked before the inner
163 circumference. The cylindrical hydrogels were cut perpendicularly into three approximately
164 equal masses and labelled as 1, 2, and 3 as shown in the photographic insert in Fig. 1. The
165 aliquot hydrogels labelled 1 and 3 were mainly composed of the outer geometry of the
166 hydrogels, while the aliquot labelled 2 was composed of the mid portion of the hydrogel.
167 Afterwards, the aliquots were dried and the polymer weight fraction, which is the ratio of dry
168 cut to swollen cut was reported.

169 **2.6 Fourier transform infrared spectroscopy (FTIR) analysis**

170 The interaction between the alginate and the two types of CNF at different loadings were
171 studied from dried samples using Spectrum 100 FTIR (Perkin Elmer, USA), in the attenuated
172 total reflectance (ATR) mode. Spectral data were collected from 4000 to 500 cm^{-1} after 8 scans.

173 **2.7 Rheological measurements of the hydrogels**

174 The viscoelastic properties of the equilibrium swollen hydrogels were analysed using a 25 mm
175 parallel plate geometry on MCR 301 rheometer (Anton Paar, Austria). A limited amplitude
176 sweep was first conducted from 0.001 % to 0.2 % shear strain at 1 Hz frequency in order to
177 determine the Linear Viscoelastic Region (LVR). This is the region where the microstructural
178 response and associated viscoelastic properties of a material are independent of the degree of
179 deformation imposed (Goodwin & Hughes, 2008). This was followed by three frequency
180 sweeps: an initial sweep from 1 Hz to 10 Hz, followed by a second sweep from 0.1 Hz to 50
181 Hz and finally a third sweep from 1 Hz to 10 Hz, all carried out at 0.01% shear strain. The
182 repeated frequency sweeps were carried out to verify whether the samples are within their linear
183 viscoelastic region during the frequency sweeps. This should appear as an overlap of the three

184 frequency sweeps. Each sample was tested in duplicates. Since the three frequency sweep
185 regimes were overlapping for all the samples, only the average of the second frequency sweep
186 (0.1 Hz to 50 Hz) is hereby reported.

187 **2.8 Mechanical measurements of the hydrogels by uniaxial compression tests**

188 The equilibrium-swollen hydrogels were carefully cut with an 8 mm diameter metal cutter in a
189 pool of deionised water just before the compression tests to prevent water loss by drying. The
190 sample height varied between 5.3 to 6.2 mm with an average diameter of 8 mm. The samples
191 were compressed to 4.3 – 5.2 mm (compression gap of 1 mm) at a velocity of 100 $\mu\text{m/s}$ using
192 an 8 mm parallel plate geometry (PP8) on MCR 301 rheometer (Anton Paar, Austria). The
193 upper platen was gently lowered until the normal force was between 0 N and 0.1 N. The normal
194 force-displacement data obtained from the rheometer was transformed to stress-deformation
195 factor data using Equations 2 and 3. The deformation factor is related to strain but considers
196 the dimensional (x, y, z) changes upon compression (Flory, 1953). The compressive modulus
197 (G) was calculated as the slope of the linear region from the plot of stress vs deformation factor
198 as shown in Equation 4. An average value of G from five compression tests is hereby reported
199 for each sample.

$$200 \quad \sigma = F/A \quad \text{Equation 2}$$

$$201 \quad \gamma = -(\lambda - \lambda^{-2}) \quad \text{Equation 3}$$

$$202 \quad G = \sigma/\gamma \quad \text{Equation 4}$$

203 Here, σ is the compressive stress, F is the normal force, $A = (\pi r^2)$ is the cross-sectional area
204 of the sample geometry, in this case a cylinder, γ is the deformation factor, and λ is the ratio of
205 the final height of sample to the initial height, known as the compression ratio.

206 Using the data from compression tests and equilibrium swelling in water, the elastically
207 effective chains of the hydrogels, ν_e (mol/cm^3) were determined. This is the chain ends that are
208 connected to the active crosslinks within the hydrogel network. This method of determining
209 the elastically effective chains is based on the affine network model, which assumes that
210 crosslinks are firmly connected to the elastic body, and elastically effective chains deform in
211 the same manner with the macroscopic body (Akagi et al., 2011). ν_e was calculated for each
212 sample using the modified version of Flory's equation for the deformation of swollen network
213 (Flory, 1953) given in Equation 5 as obtained from Muniz & Geuskens (2001).

214
$$v_e = G / RT(\phi_r/\phi_s)^{2/3}\phi_s$$
 Equation 5

215 Here, G is the compressive modulus in Pa, R is the universal gas constant ($8.314 \times 10^6 \text{ cm}^3 \text{ Pa}$
216 $\text{mol}^{-1} \text{ K}^{-1}$), T is the absolute temperature (293 K), ϕ_r is the volume fraction of the polymer in
217 the relaxed state, i.e., the hydrogel formed in the agar mould after the excess CaCl_2 has been
218 leached and ϕ_s is the volume fraction of polymer at equilibrium swollen state. The volume of
219 SA, CNF and water were converted from their weights using the density of SA (1.6 g/cm^3),
220 cellulose (1.5 g/cm^3) and water at $20 \text{ }^\circ\text{C}$ (0.9982 g/cm^3).

221 **2.9 Morphological assessment**

222 Morphological investigations of the cross-sections of freeze-dried SA, SA_MCNF_5 and
223 SA_CMCNF_5 were carried out using a field emission scanning electron microscope (FE-
224 SEM), ZEISS SUPRA 40 VP. The freeze-dried equilibrium swollen hydrogels allow the
225 probing of the network structure created within the hydrogel during the crosslinking process.
226 Prior to image analysis, all samples were obtained by fragile fractures in liquid nitrogen and
227 were thinly sputter-coated with carbon using Polaron E5100 sputter coater. The pore sizes of
228 the hydrogels were measured from the FE-SEM images using image analysis software
229 (ImageJ).

230 **2.10 Statistical analysis**

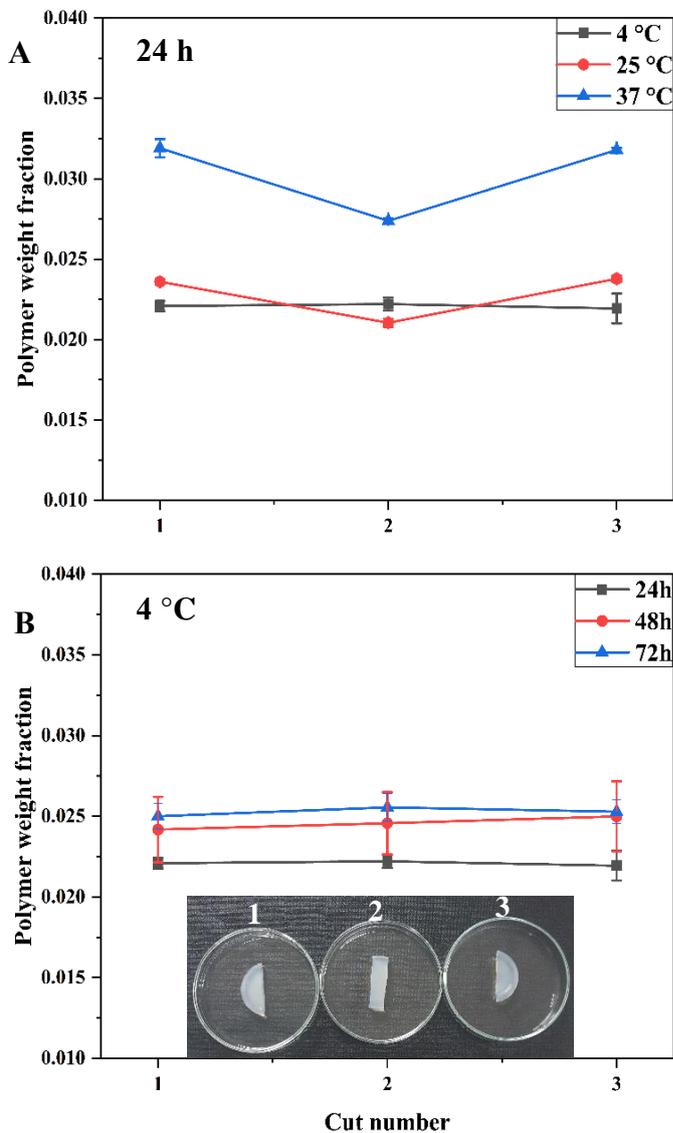
231 Results are presented as means \pm standard deviation. Each loading of either MCNF or CMCNF
232 was compared to the alginate using two-sample t-test on OriginPro 2018 software. The
233 difference between means is considered significant when $p < 0.05$.

234 **3. Results and Discussion**

235 **3.1 Gelation and Homogeneity**

236 In this study, CaCl_2 enriched agar gels were used to control the gelation of SA by allowing
237 slower diffusion of the calcium ions from the agar gels into the SA solutions. Visual
238 observations of the hydrogels showed that the crosslinking process initially began from the
239 outer circumference of the alginate solution in contact with the agar mould, before gradually
240 diffusing into the inner circumference of the solution. The effect of gelation temperature and
241 the duration of crosslinking on the homogeneity of the neat SA hydrogels can be seen from

242 Fig. 1A (crosslinked for 24 h at 4 °C, 25 °C and 37°C) and Fig. 1B (crosslinked at 4°C for 24
243 h, 48 h and 72 h).



244

245 **Fig. 1:** Effect of temperature (A) and time (B) on the homogeneity of SA hydrogels.
246 Photographic insert of the equilibrium swollen hydrogel cuts

247 There was no significant change in the polymer weight fractions across the three cuts of the
248 hydrogels prepared at 4 °C as shown in Fig. 1A, indicating a homogeneous network formation
249 within the gels. This can be attributed to the slower diffusion of the calcium ions from the agar
250 gels into the SA solution at 4 °C. These hydrogels also had reduced shrinkage at the end of the
251 crosslinking process. However, when the SA solution was crosslinked at 25 °C and 37 °C,
252 heterogeneous hydrogels resulted, with the mid-point of the hydrogels (cut number 2) having
253 a lower polymer fraction compared to the outer cuts. The reduction in polymer fraction at the

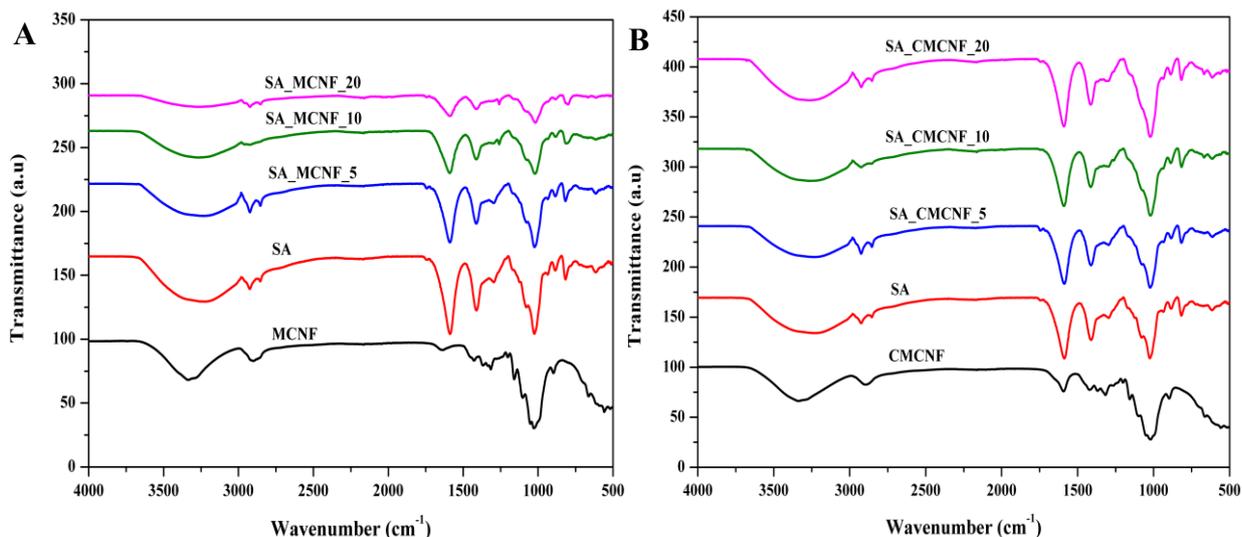
254 midpoint of the hydrogels was very significant for the hydrogels prepared at 37 °C, directly
255 implying a higher water content and reduced crosslinking density at the mid-point of the
256 hydrogel. These results and observations show that the temperature of gelation plays a major
257 role on the homogeneity of these hydrogels when using the agar gel moulds.

258 The various durations of gelation at 4 °C do not have any significant effect on the
259 polymer weight fractions across the three cuts as shown in Fig. 1B, implying that the duration
260 of crosslinking does not affect the homogeneity of the hydrogels. Henceforth, the composite
261 hydrogels reported herein are for those crosslinked at 4 °C for a 24 h period.

262 **3.2 Surface interaction from FTIR analysis**

263 FTIR analysis was carried out to probe the interaction between the sodium alginate and the two
264 types of CNF upon crosslinking. The overlay of the FTIR spectrum for SA_MCNF and
265 SA_CMCNF composite hydrogels are shown in Fig. 2A and 2B respectively. All the spectrum
266 from the composite hydrogels presented the same peaks as the alginate matrix. The broad peaks
267 identified in all the spectrum between 3227-3335 cm⁻¹ were assigned to the O-H stretching
268 vibrations, while the peaks identified between 2924 - 2902 cm⁻¹, 1590 - 1587 cm⁻¹, 1414 - 1411
269 cm⁻¹ and 1024 -1015 cm⁻¹ were assigned to the C-H, COO⁻ (asymmetric), COO⁻ (symmetric)
270 and C-O stretching vibrations respectively (Sirviö, Kolehmainen, Liimatainen, Niinimäki, &
271 Hormi, 2014).

272 A shift in the O-H stretching vibration to higher wavenumber was seen for
273 SA_MCNF_10 and SA_MCNF_20 (from 3227 to 3264 cm⁻¹) and for SA_CMCNF_10 and
274 SA_CMCNF_20 (from 3227 to 3266 cm⁻¹), indicating an increase in the O-H functional
275 groups, as the percentage loadings of both CNFs increases. There were significant shifts of the
276 C-O peak from 1024 cm⁻¹ to 1020 cm⁻¹, and 1024 cm⁻¹ to 1017 cm⁻¹ for SA_MCNF_10 and
277 SA_MCNF_20 respectively. A similar shift to lower wavenumber of the C-O peak was seen
278 with alginate film having 50 % microfibrillated cellulose and was attributed to an interaction
279 between alginate and residual hemicellulose (Sirviö et al., 2014). However, the conclusion of
280 the authors cannot be adopted in this case as the cellulose material used for the preparation of
281 the MCNF was from dissolving pulp, having negligible hemicellulose content. These shifts
282 toward lower wavenumbers at these percentage loadings imply an interaction between alginate
283 and MCNF at lower bond energy in comparison with the pure alginate.



284

285 **Fig. 2:** FTIR spectrum overlay of SA_MCNF and SA_CMCNF composite hydrogels with each
 286 of the starting materials

287 It was of interest to investigate whether the carboxymethyl groups of CMCNF at the
 288 amounts studied interacted with the alginate in such a way as to contribute to ionic crosslinking
 289 as has been reported for carboxyl groups of TEMPO oxidised CNF (Lin et al., 2012). This
 290 would be observed as a shift in the asymmetric and symmetric peaks of COO⁻ groups. There
 291 was only a 3 cm⁻¹ shift to higher wavenumber for SA_CMCNF_20, which is considered
 292 statistically significant. Therefore, it can be assumed that there is an interaction between
 293 alginate and CMCNF at the COO⁻ functional group for SA_CMCNF_20, which may have some
 294 minor contribution to ionic crosslinking. There was no significant shift in the peaks of all other
 295 observable functional groups of alginates with the addition of CMCNF at different percentage
 296 loadings.

297 3.3 Effect of CNF loadings on the swelling of never-dried hydrogels in water

298 The amount of water imbibed by a three-dimensional network alginate hydrogel system would
 299 depend on the amount of free hydrophilic hydroxyl groups which are not participating in ionic
 300 crosslinking and on the degree of crosslinking, in other words, the void spaces created within
 301 the hydrogels after crosslinking (Draget, Skjåk-Bræk, & Smidsrød, 1997). The effects of
 302 varying loadings of the CNF materials on the swelling degree of SA hydrogels from never-
 303 dried samples were studied by swelling the hydrogels to an equilibrium state in water and the
 304 results presented in Table 1.

305 **Table 1:** Swelling degree, storage modulus and crosslinking degree parameters of the alginate-based
 306 composite hydrogels.

Sample	Swelling degree	Storage modulus, G' (kPa) @ 1 Hz	ν_e (mol/cm ³)
SA	42.6 ±0.8	44.5 ±0.0	6.9E-4 ±2.0E-5
SA_MCNF_5	38.0 ±1.9	56.4 ±4.5	8.5E-4 ±1.9E-5
SA_MCNF_10	40.3 ±1.5	54.6 ±4.8	7.8E-4 ±1.4E-5
SA_MCNF_20	39.2 ±0.4	56.4 ±9.5	5.5E-4 ±1.6E-5
SA_CMCNF_5	44.8 ±0.5	48.9 ±3.4	7.9E-4 ±3.7E-5
SA_CMCNF_10	45.8 ±0.1	50.4 ±0.5	6.8E-4 ±4.6E-5
SA_CMCNF_20	42.5 ±2.2	64.7 ±9.9	6.9E-4 ±7.6E-5

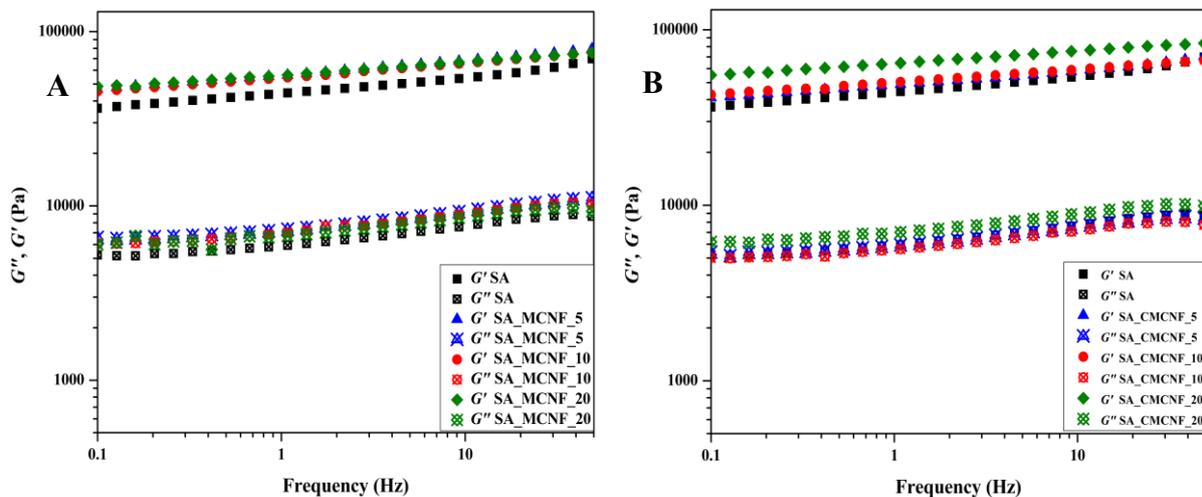
307

308 The addition of MCNF at all the percentage loadings studied led to an overall
 309 significant reduction in the swelling degree of SA_MCNF hydrogels in comparison to the neat
 310 SA hydrogel. The least swelling degree was observed for SA_MCNF_5 and 11 % lower than
 311 neat SA hydrogel. The reduction in swelling was also reported by Huq et al. (2012) with the
 312 addition of 5 wt. % cellulose nanocrystals to sodium alginate films and was attributed to an
 313 increase in mechanical property.

314 On the other hand, the swelling degree of SA_CMCNF_5 and SA_CMCNF_10 were
 315 significantly improved when compared to the neat SA, whereas no change in swelling degree
 316 was observed for SA_CMCNF_20. Similar increase in aqueous swelling behaviour was
 317 observed by Lin et al., (2014) when increasing the amount of carboxymethylated bacterial
 318 cellulose in alginate-based hydrogels. The increase in swelling was attributed to the presence
 319 of the anionic hydrophilic carboxymethyl groups on the surface of CMCNF. The anionic
 320 groups which are not contributing to ionic crosslinking bring about electrostatic repulsive
 321 forces on the fibre surface, which promotes fibre swelling, and adds to the overall swelling
 322 degree of the hydrogels. These anionic groups are however not present on the surface MCNF
 323 and do not contribute to the swelling, hence the observed reduced swelling.

324 **3.4 Effect of the CNF materials on the viscoelastic properties of the alginate-based**
325 **hydrogels**

326 The G' of the hydrogels at 1 Hz are presented in Table 1. In addition, the overlay of the
327 frequency sweeps of the SA_MCNF and SA_CMCNF composite hydrogels at a constant strain
328 value of 0.01% are shown in Fig. 3. All the composite hydrogels studied show a higher storage
329 modulus (G') than loss modulus (G''), an indication of complete gelled network material
330 (Magami, 2017). In addition, the G' of all the hydrogels are almost independent of the
331 frequency applied which is also a prevalent characteristic of a gelled network system.

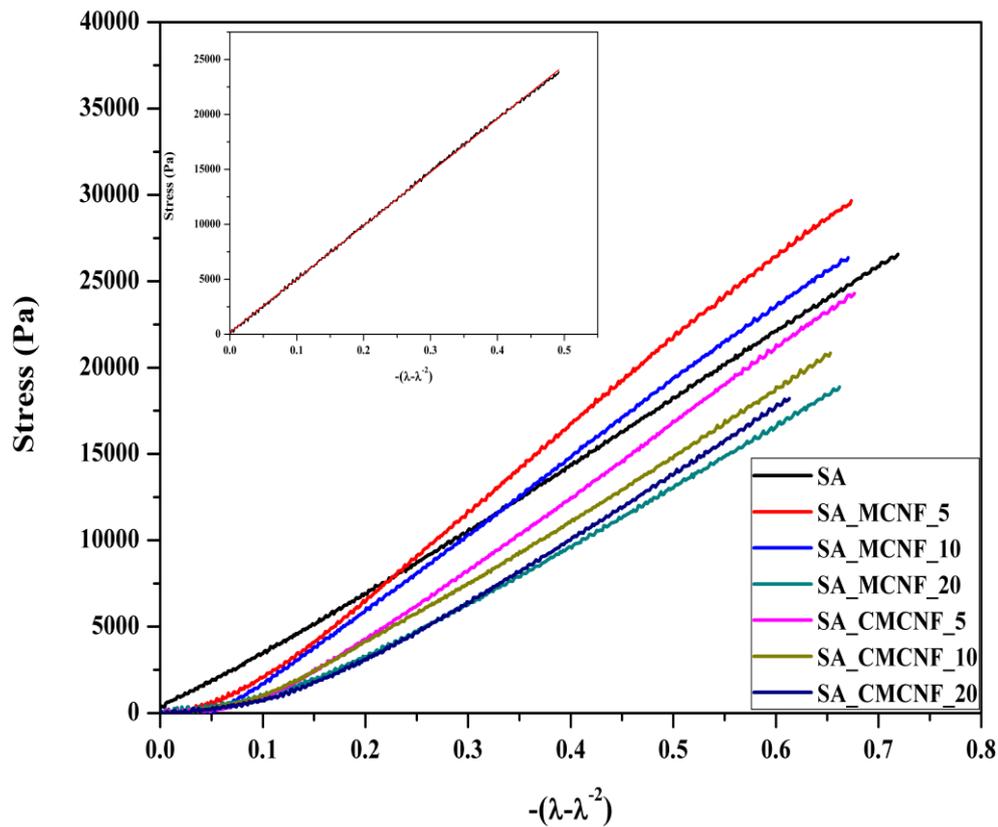


332
333 **Fig. 3:** Overlay of frequency sweeps of SA_MCNF (A) and SA_CMCNF (B) hydrogels

334 The addition of 5 wt. % MCNF led to a 27 % increase in G' compared to the neat SA
335 hydrogel, further increases in the percentage loading of MCNF did not result in a significant
336 effect on the storage modulus of the hydrogels. On the other hand, the addition of 5 wt. %
337 CMCNF led to a 10 % increase in G' compared to the neat SA hydrogel. The higher storage
338 modulus of SA_MCNF_5 in comparison to the SA_CMCNF_5 suggests a higher reinforcing
339 ability of MCNF compared to CMCNF. The higher storage modulus could also explain the low
340 swelling degree of SA_MCNF composite hydrogels. Although the increase in G' was lower
341 for SA_CMCNF_5, there was a linear and significant increase in the storage modulus with
342 further increases in the amounts CMCNF. The significant increase in G' with the addition of
343 10 wt. % and 20 wt. % CMCNF can be attributed to the presence of the anionic carboxymethyl
344 groups which have the potentials to contribute to the gel-like behaviour of alginate, given that
345 carboxymethylated cellulose (CMC) is a well-known rheology modifier.

346 **3.5 Effect of CNF materials on the compressive modulus and elastically effective**
347 **chains of alginate-based hydrogels**

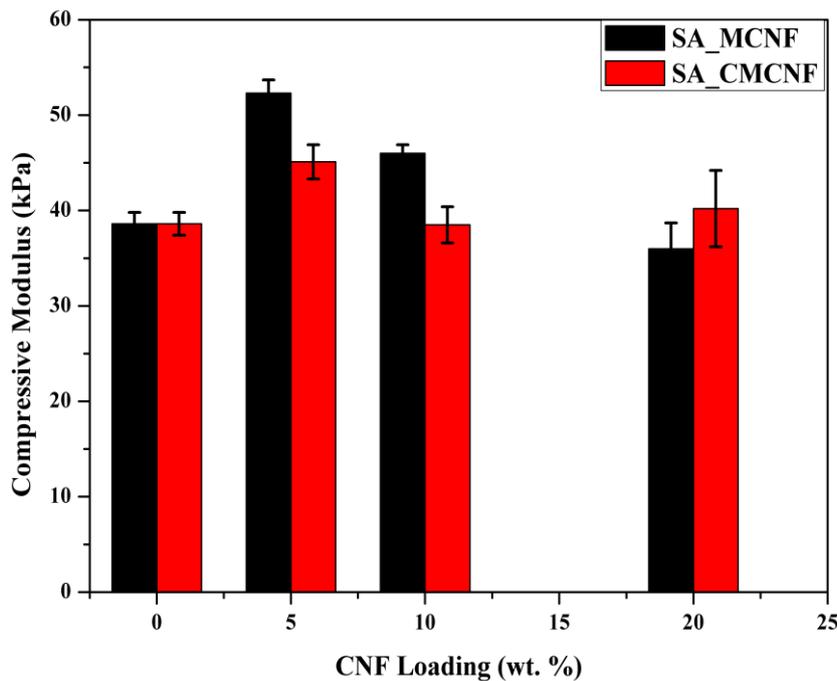
348 The mechanical properties of the hydrogels were further characterised by carrying out
349 compression tests on the equilibrium-swollen hydrogels. An overlay of the plot of stress vs
350 deformation factor for all the hydrogel is shown in Fig. 4.



351
352 **Fig. 4:** Stress-deformation factor relationship of all hydrogels with a representative insert of
353 the corrected plot without a toe region.

354 All hydrogels tested showed a toe region at the beginning of the test, up to 10 % deformation,
355 before showing a linear relationship between stress and deformation factor. The toe region
356 manifests because of imperfect hydrogel surface as the upper platen of the rheometer makes
357 contact with the entire area of the hydrogel cylinder (Muniz & Geuskens, 2001). It was
358 observed that 0.1 N normal force was enough to make good contact with the hydrogels.
359 Subsequently, the data set was corrected by subtracting the value of stress and deformation
360 factor at 0.1 N from all data set, to obtain the plot shown as the insert image (representative
361 image) on Fig. 4. The linear fit of the plot was then carried out using OriginPro 2018 software
362 and the slope of the plot reported as the compressive modulus (G).

363 The compressive modulus of SA, SA_MCNF and SA_CMCNF hydrogels are shown
 364 in Fig. 5. The hydrogel composite materials showed highest compressive modulus at 5 wt. %
 365 MCNF and CMCNF loadings. The addition of 5 wt. % MCNF resulted in a 36 % increase in
 366 compressive modulus of the hydrogels in comparison with the neat SA hydrogel, while a 17 %
 367 increase in compressive modulus was observed with the addition of CMCNF at the same
 368 percentage loading. This result confirms that MCNF is more efficient in reinforcing SA than
 369 CMCNF.



370

371 **Fig. 5:** Compressive modulus of SA_MCNF and SA_CMCNF hydrogels

372 The SA_MCNF_10 and SA_MCNF_20 hydrogels with lower swelling degree and
 373 higher storage modulus are also expected to have higher compressive modulus in comparison
 374 to the neat SA hydrogels. However, lower compressive moduli were seen for these samples. It
 375 should be noted that at 10 wt. % (0.002 wt. fraction) and 20 wt. % (0.004 wt. fraction) MCNF
 376 and CMCNF loadings, the fibrils are beyond the respective connectivity threshold of 0.0014
 377 wt. fraction and 0.00065 wt. fraction (Onyianta et al., 2018). Therefore, there is a tendency for
 378 the fibrils to form aggregates within the hydrogels, leading to network imperfection. The
 379 compressive deformation imposed on the hydrogels appears to be more sensitive to these
 380 fibrillar aggregation occurring at higher percentage loading of the CNF materials. These
 381 fibrillar aggregations and entanglements seems to have offset any reinforcing capacity accrued
 382 at lower percentage loading. The rotational shearing force imposed during the frequency
 383 sweeps might not have identified the presence of fibril kinks and aggregation but able to detect

384 the increase in the solid-like component of the hydrogels. Hence, the G' is seen to plateau or
385 increase linearly when the respective amount of MCNF and CMCNF is increased.

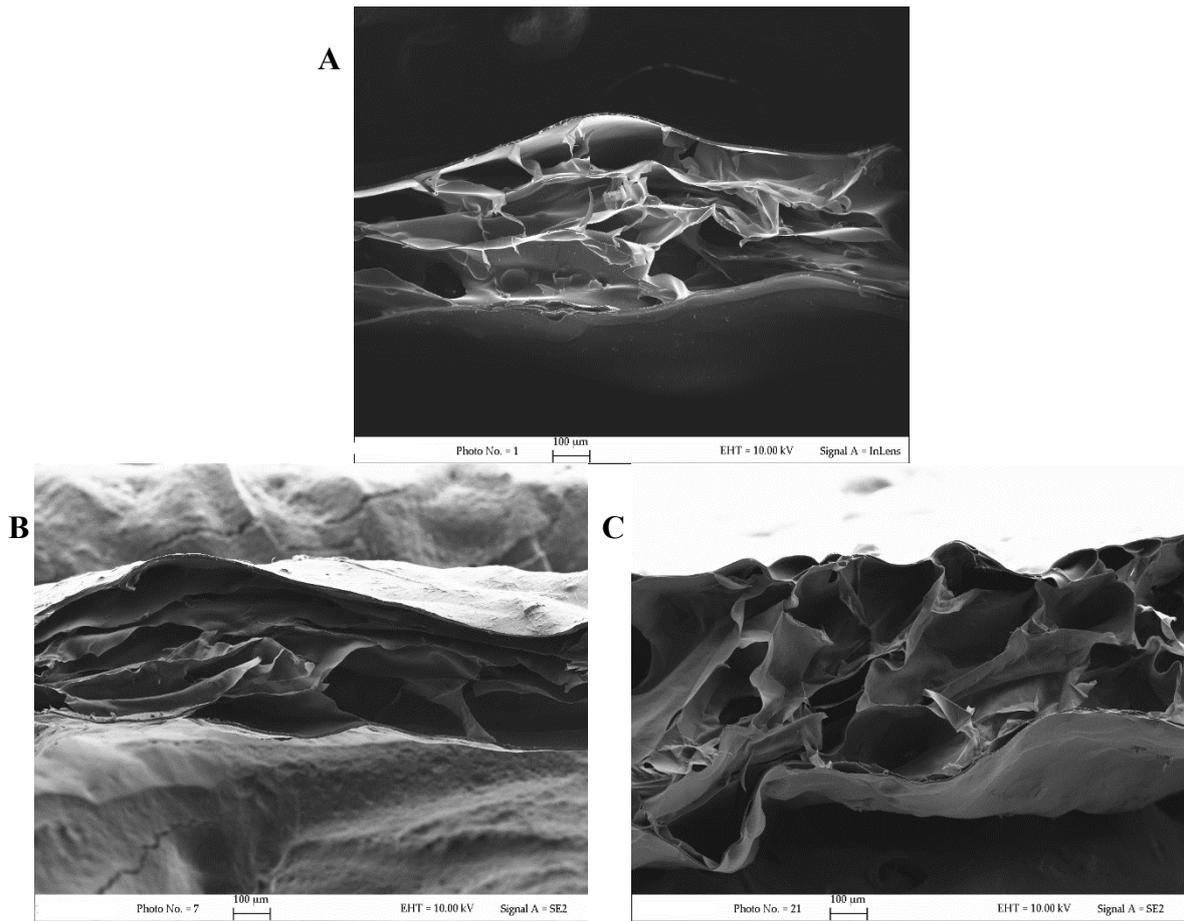
386 To further elucidate the effect of the CNF materials on SA hydrogels' network
387 structure, the modified Flory's equation for rubber elasticity was used to calculate the moles
388 per cm^3 of the elastically effective chains within the SA_CNF composite hydrogels. This
389 equation basically relates the compressive modulus with the inverse of equilibrium swelling
390 degree as shown in Equation 5. The effect of the CNF type and loading on the moles of
391 elastically effective chains of the hydrogels is shown in Table 1. An increase in the elastically
392 effective chains of SA_MCNF_5 and SA_MCNF_10 when compared to the neat alginate was
393 observed. This increase is an indication of the formation of physical networks, through
394 intermolecular interactions. However, at 20 wt. % MCNF, aggregation of fibrils begins to
395 occur, resulting in the formation of imperfections, hence the low ν_e .

396 Also, there was an increase in ν_e for SA_CMCNF_5, however, there was no significant
397 change in the elastically effective chains of SA_CMCNF_10 and SA_CMCNF_20. Although
398 the FTIR data analysis suggested a possible minor contribution in ionic crosslinking for
399 SA_CMCNF_20, these crosslinks if present, may not have been elastically effective. Thereby
400 explaining the insignificant change in the compressive modulus of the sample.

401 **3.6 Morphological properties of SA_CNF composite hydrogels.**

402 The hydrogel with the highest compressive modulus for each CNF type was probed to identify
403 the degree and nature of pores created upon equilibrium swelling in water and freeze-drying.
404 The cross-sectional images of SA, SA_MCNF_5 and SA_CMCNF_5 samples are shown in
405 Fig. 6, having a respective average porous size of $114 \pm 46 \mu\text{m}$, $84 \pm 43 \mu\text{m}$ and $157 \pm 86 \mu\text{m}$.

406 The incorporation of MCNF into the SA hydrogel resulted in a rather compact and
407 distinct layered structure, having lower average pore size than the neat SA hydrogel and the
408 SA_CMCNF_5 hydrogel, which both have network structures that are more porous. The
409 surface repulsive carboxymethyl groups alongside the carboxyl groups on SA would result in
410 a net increased void spaces within the polymer, leading to larger intake of water. Whereas the
411 attractive hydroxyl groups on the surface of the MCNF would lead to an increased inter and
412 intramolecular attraction of the fibrils within the SA_MCNF_5 composite, hence the observed
413 lower porosity relative to the neat SA hydrogel. This explains the reduced water uptake of
414 SA_MCNF_5 hydrogel when compared to SA and SA_CMCNF_5 hydrogels as shown in
415 Table 1.



416

417 **Fig. 6:** Cross-sectional FE-SEM images of SA hydrogel (A), SA_MCNF_5 (B) and
 418 SA_CMCNF_5 (C)

419 **Conclusions**

420 The use of calcium chloride enriched agar gel moulds at 4 °C resulted in the formation of
 421 homogeneously crosslinked alginate. The addition of 5 wt. % MCNF led to a 36 % increase in
 422 compressive modulus, an increase in the elastically effective chains and storage modulus. In
 423 addition, the SA_MCNF_5 composite hydrogel had less porous structure, because of the lack
 424 of anionic repulsive groups on the surface of MCNF, which contributed to the observed reduced
 425 swelling in water. On the other hand, the presence of anionic carboxymethyl groups on the
 426 surface of the CMCNF led to a slightly improved swelling degree, a more porous network
 427 structure, having smaller increases in the compressive modulus, elastically effective chains and
 428 storage modulus when compared to the neat alginate hydrogel. These positive effects were
 429 observed at lower percentage loadings of the CNFs (5 wt. %). Increasing the percentage
 430 loadings however did not lead to improvement in material properties of the hydrogels, due to
 431 possible fibril aggregation. Therefore, only 5 wt. % CNF materials are required to prepare

432 alginate hydrogels that could find applications in areas where good swelling and mechanical
433 strength are needed.

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438 **References**

- 439 Abdul Khalil, H. P. S., Davoudpour, Y., Islam, M. N., Mustapha, A., Sudesh, K., Dungani, R., & Jawaid, M.
440 (2014). Production and modification of nanofibrillated cellulose using various mechanical processes: A
441 review. *Carbohydrate Polymers*, 99, 649–665. <https://doi.org/10.1016/j.carbpol.2013.08.069>
- 442 Ahmed, E. M. (2015). Hydrogel: Preparation, characterization, and applications: A review. *Journal of Advanced*
443 *Research*, 6(2), 105–121. <https://doi.org/10.1016/j.jare.2013.07.006>
- 444 Akagi, Y., Katashima, T., Katsumoto, Y., Fujii, K., Matsunaga, T., Chung, U. Il, ... Sakai, T. (2011).
445 Examination of the theories of rubber elasticity using an ideal polymer network. *Macromolecules*.
446 <https://doi.org/10.1021/ma201088r>
- 447 Aravamudhan, A., Ramos, D. M., Nada, A. A., & Kumbar, S. G. (2014). Chapter 4 - Natural Polymers:
448 Polysaccharides and Their Derivatives for Biomedical Applications. In M. Kumbar, Sangamesh G.;
449 Laurencin, Cato T.; Deng (Ed.), *Natural and Synthetic Biomedical Polymers* (pp. 67–89). Elsevier.
450 <https://doi.org/https://doi.org/10.1016/B978-0-12-396983-5.00004-1>
- 451 Augst, A. D., Kong, H. J., & Mooney, D. J. (2006). Alginate hydrogels as biomaterials. *Macromolecular*
452 *Bioscience*, 6(8), 623–633. <https://doi.org/10.1002/mabi.200600069>
- 453 Aulin, C., Johansson, E., Wågberg, L., & Lindström, T. (2010). Self-organized films from cellulose i nanofibrils
454 using the layer-by-layer technique. *Biomacromolecules*, 11(4), 872–882.
455 <https://doi.org/10.1021/bm100075e>
- 456 Bajpai, M., Shukla, P., & Bajpai, S. K. (2016). Ca(II)+Ba(II) ions crosslinked alginate gels prepared by a novel
457 diffusion through dialysis tube (DTDT) approach and preliminary BSA release study. *Polymer*
458 *Degradation and Stability*. <https://doi.org/10.1016/j.polymdegradstab.2016.09.027>
- 459 Beyene, D., Chae, M., Dai, J., Danumah, C., Tosto, F., Demesa, A. G., & Bressler, D. C. (2017). Enzymatically-
460 Mediated Co-Production of Cellulose Nanocrystals and Fermentable Sugars. *Catalysts* .
461 <https://doi.org/10.3390/catal7110322>
- 462 Cardoso, M. J., Costa, R. R., & Mano, J. F. (2016). Marine origin polysaccharides in drug delivery systems.
463 *Marine Drugs*, 14(2), 1–27. <https://doi.org/10.3390/md14020034>

464 Draget, K. I., Skjåk-Bræk, G., & Smidsrød, O. (1997). Alginate based new materials. *International Journal of*
465 *Biological Macromolecules*, 21(1–2), 47–55. [https://doi.org/10.1016/S0141-8130\(97\)00040-8](https://doi.org/10.1016/S0141-8130(97)00040-8)

466 Flory, P. J. (1953). Chapter XI: Rubber Elasticity. In P. J. Flory (Ed.), *Principles of Polymer Chemistry* (pp.
467 432–493).

468 Goodwin, J. W., & Hughes, R. W. (2008). *Rheology for Chemists*. Royal Society of Chemistry.
469 <https://doi.org/10.1039/9781847558046>

470 Guilherme, M. R., Aouada, F. A., Fajardo, A. R., Martins, A. F., Paulino, A. T., Davi, M. F. T., ... Muniz, E. C.
471 (2015). Superabsorbent hydrogels based on polysaccharides for application in agriculture as soil
472 conditioner and nutrient carrier: A review. *European Polymer Journal*, 72, 365–385.
473 <https://doi.org/10.1016/j.eurpolymj.2015.04.017>

474 Hubbe, M. A., Ferrer, A., Tyagi, P., Yin, Y., & Salas, C. (2017). com Nanocellulose in Thin Films, Coatings,
475 and Plies for Packaging Applications: A Review, 12, 2143–2233.

476 Huq, T., Frascini, C., Khan, A., Riedl, B., Bouchard, J., & Lacroix, M. (2017). Alginate based nanocomposite
477 for microencapsulation of probiotic: Effect of cellulose nanocrystal (CNC) and lecithin. *Carbohydrate*
478 *Polymers*, 168, 61–69. <https://doi.org/10.1016/j.carbpol.2017.03.032>

479 Huq, T., Salmieri, S., Khan, A., Khan, R. A., Le Tien, C., Riedl, B., ... Lacroix, M. (2012). Nanocrystalline
480 cellulose (NCC) reinforced alginate based biodegradable nanocomposite film. *Carbohydrate Polymers*,
481 90(4), 1757–1763. <https://doi.org/10.1016/j.carbpol.2012.07.065>

482 Isogai, A., Saito, T., & Fukuzumi, H. (2011). TEMPO-oxidized cellulose nanofibers. *Nanoscale*, 3(1), 71–85.
483 <https://doi.org/10.1039/C0NR00583E>

484 Jonoobi, M., Oladi, R., Davoudpour, Y., Oksman, K., Dufresne, A., Hamzeh, Y., & Davoodi, R. (2015).
485 Different preparation methods and properties of nanostructured cellulose from various natural resources
486 and residues: a review. *Cellulose*, 22(2), 935–969. <https://doi.org/10.1007/s10570-015-0551-0>

487 Klemm, D., Kramer, F., Moritz, S., Lindström, T., Ankerfors, M., Gray, D., & Dorris, A. (2011).
488 Nanocelluloses: A new family of nature-based materials. *Angewandte Chemie - International Edition*,
489 50(24), 5438–5466. <https://doi.org/10.1002/anie.201001273>

490 Kong, H. J., & Mooney, D. J. (2003). The Effects of Poly(Ethyleneimine) (PEI) Molecular Weight on
491 Reinforcement of Alginate Hydrogels. *Cell Transplantation*.
492 <https://doi.org/10.3727/000000003108747253>

493 Kuo, C. K., & Ma, P. X. (2001). Ionically crosslinked alginate hydrogels as scaffolds for tissue engineering: Part
494 1. Structure, gelation rate and mechanical properties. *Biomaterials*, 22(6), 511–521.
495 [https://doi.org/10.1016/S0142-9612\(00\)00201-5](https://doi.org/10.1016/S0142-9612(00)00201-5)

496 Lee, S. B., Seo, S. M., Lim, Y. M., Cho, S. K., Lee, Y. M., & Nho, Y. C. (2004). Preparation of alginate/poly(N-
497 isopropylacrylamide) hydrogels using gamma-ray irradiation grafting. *Macromolecular Research*, 12(3),
498 269–275. <https://doi.org/10.1007/BF03218399>

499 Leppiniemi, J., Lahtinen, P., Paajanen, A., Mahlberg, R., Metsä-Kortelainen, S., Pinomaa, T., ... Hytönen, V. P.
500 (2017). 3D-Printable Bioactivated Nanocellulose-Alginate Hydrogels. *ACS Applied Materials and*
501 *Interfaces*, 9(26), 21959–21970. <https://doi.org/10.1021/acsami.7b02756>

502 Lin, N., Bruzzese, C., & Dufresne, A. (2012). TEMPO-oxidized nanocellulose participating as crosslinking aid
503 for alginate-based sponges. *ACS Applied Materials and Interfaces*, 4(9), 4948–4959.
504 <https://doi.org/10.1021/am301325r>

505 Lin, Q., Zheng, Y., Ren, L., Wu, J., Wang, H., An, J., & Fan, W. (2014). Preparation and characteristic of a
506 sodium alginate/carboxymethylated bacterial cellulose composite with a crosslinking semi-
507 interpenetrating network. *Journal of Applied Polymer Science*, 131(3), 1–9.
508 <https://doi.org/10.1002/app.39848>

509 Ma, X., Li, R., Zhao, X., Ji, Q., Xing, Y., Sunarso, J., & Xia, Y. (2017). Biopolymer composite fibres composed
510 of calcium alginate reinforced with nanocrystalline cellulose. *Composites Part A: Applied Science and*
511 *Manufacturing*, 96, 155–163. <https://doi.org/10.1016/j.compositesa.2017.02.021>

512 Magami, S. M. (2017). In situ viscoelasticity and in situ thermo- responsiveness in acrylic acid-based soft
513 hydrogels In situ viscoelasticity and in situ thermo-responsiveness in acrylic acid-based soft hydrogels. In
514 *IOP Conference Series: material Science and Engineering*. [https://doi.org/10.1088/1757-](https://doi.org/10.1088/1757-899X/264/1/012019)
515 [899X/264/1/012019](https://doi.org/10.1088/1757-899X/264/1/012019)

516 Muniz, E. C., & Geuskens, G. (2001). Compressive elastic modulus of polyacrylamide hydrogels and semi-IPNs
517 with poly(N-isopropylacrylamide). *Macromolecules*, 34(13), 4480–4484.
518 <https://doi.org/10.1021/ma001192l>

519 Onyianta, A. J., Dorris, M., & Williams, R. L. (2018). Aqueous morpholine pre-treatment in cellulose nanofibril
520 (CNF) production: comparison with carboxymethylation and TEMPO oxidation pre-treatment methods.
521 *Cellulose*, 25(2), 1047–1064. <https://doi.org/10.1007/s10570-017-1631-0>

522 Park, S. Bin, Lih, E., Park, K. S., Joung, Y. K., & Han, D. K. (2017). Biopolymer-based functional composites
523 for medical applications. *Progress in Polymer Science*, 68, 77–105.
524 <https://doi.org/10.1016/j.progpolymsci.2016.12.003>

525 Sannino, A., Pappadà, S., Madaghiele, M., Maffezzoli, A., Ambrosio, L., & Nicolais, L. (2005). Crosslinking of
526 cellulose derivatives and hyaluronic acid with water-soluble carbodiimide. *Polymer*.
527 <https://doi.org/10.1016/j.polymer.2005.10.048>

528 Sirviö, J. A., Kolehmainen, A., Liimatainen, H., Niinimäki, J., & Hormi, O. E. O. (2014). Biocomposite
529 cellulose-alginate films: Promising packaging materials. *Food Chemistry*, 151, 343–351.
530 <https://doi.org/10.1016/j.foodchem.2013.11.037>

531 Smyth, M., M'Bengue, M. S., Terrien, M., Picart, C., Bras, J., & Foster, E. J. (2018). The effect of hydration on
532 the material and mechanical properties of cellulose nanocrystal-alginate composites. *Carbohydrate*
533 *Polymers*, 179(September 2017), 186–195. <https://doi.org/10.1016/j.carbpol.2017.09.002>

- 534 Stagnaro, P., Schizzi, I., Utzeri, R., Marsano, E., & Castellano, M. (2018). Alginate-polymethacrylate hybrid
535 hydrogels for potential osteochondral tissue regeneration. *Carbohydrate Polymers*.
536 <https://doi.org/10.1016/j.carbpol.2018.01.012>
- 537 Vicini, S., Castellano, M., Mauri, M., & Marsano, E. (2015). Gelling process for sodium alginate: New technical
538 approach by using calcium rich micro-spheres. *Carbohydrate Polymers*, *134*, 767–774.
539 <https://doi.org/10.1016/j.carbpol.2015.08.064>
- 540 Vicini, S., Mauri, M., Wichert, J., & Castellano, M. (2017). Alginate gelling process: Use of bivalent ions rich
541 microspheres. *Polymer Engineering and Science*. <https://doi.org/10.1002/pen.24552>
- 542 Wågberg, L., Decher, G., Norgren, M., Lindström, T., Ankerfors, M., & Axnäs, K. (2008). The builds of
543 microfibrillated cellulose and cationic polyelectr-up of polyelectrolyte multilayeolytes. *Langmuir*, *24*(3),
544 784–795. <https://doi.org/10.1021/la702481v>
- 545